

Syntheses, Antifeedant Activity, and QSAR Analysis of New Oxa(thia)diazolyl 3(2*H*)-Pyridazinones

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Twenty-eight new oxa(thia)diazolyl 3(2*H*)-pyridazinone derivatives were synthesized. Some compounds showed good chronic growth activities against the armyworm, *Pseudaletia separata* (Walker). Their EC₅₀ values were determined in vivo. Nineteen 2-*tert*-butyl-4-chloro-5-[5'-aryl-2'-(1',3',4'-oxadiazolyl)methoxy]-3(2*H*)-pyridazinones were quantitatively analyzed using the QSAR procedure. The partial least squares method was applied to find relationships between activity and the physicochemical data. The results showed that dipole moment, molar refractivity, and log *P* are identified as critical parameters for chronic growth effects.

KEYWORDS: Syntheses; oxa(thia)diazolyl 3(2*H*)-pyridazinones; chronic growth activity; QSAR

INTRODUCTION

There has been much interest recently in natural products that exhibit antifeedant activity from an agricultural point of view (1, 2). The best known example is azadirachtin, a potent antifeedant and insect growth regulator isolated from the seed of the Indian neem tree (3, 4). However, their translation into a commercial product has been slow, partly because of the structural complexity, instability, and synthetic inaccessibility of those natural products.

In the course of screening compounds for novel and structurally simple antifeedants, we found some new oxadiazolyl 3(2*H*)-pyridazinones that showed chronic growth activity against the Asiatic corn borer, *Ostrinia furnacalis* (Guenee). The characteristic effects of the compounds to Asiatic corn borer were similar to those of azadirachtin. These data suggested that the oxadiazolyl 3(2*H*)-pyridazinones might be a new type of antifeedant (5–7).

Early work in this series concentrated on preparing less complex analogues of **IIIa** (**IIIb–t**, X¹ = O or S, X² = O, R¹ = *tert*-butyl, R² = substituted phenyl). In this paper, chemical transformations were conducted to modify three chemical moieties present in oxadiazolyl 3(2*H*)-pyridazinone and to synthesize four series of key derivatives. The structural modifications were made at the X², R¹, and R² positions of the skeletal structure shown in **Figure 1** and **Table 1** (**IVa–h**, **Va–h**, **VIa–c**, and **VIIa–f**).

Because the 1,3,4-thiadiazoles are bioisosteric analogues of the 1,3,4-oxadiazoles, which have chemical, physical, and

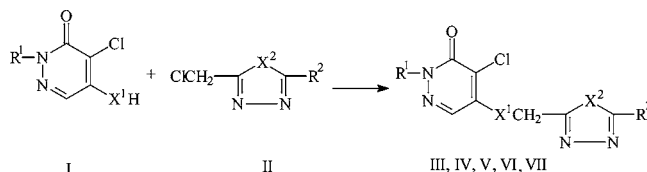


Figure 1. Synthesis of the final compounds.

biological similarities (8), some thiadiazolyl 3(2*H*)-pyridazinones (**IVa–h**, X² = S) were synthesized to elevate their activity.

The introduction of a fluorine, polyfluorine, or trifluoromethyl group into organic molecules frequently results in compounds that display more potent activity than the parent (9). Recently, polyfluorinated organic compounds have been widely employed in pharmaceuticals, agrochemicals, and new materials (10, 11). In pesticide chemistry, several commercial polyfluorinated products with their unique properties have been manufactured and used worldwide. For example, Tefluthrin is a kind of synthetic pyrethroid that has been shown to bind to the sodium channels in insects and cause knockdown and death (12), and Flufenoxuron, an insect growth regulator (13), can interfere with chitin biosynthesis in insects. In our previous research, we reported that the monofluorinated oxadiazolyl pyridazinones (**IIIf**, **IIIj**, and **IIIk**) have potent chronic growth activities against the Asiatic corn borer. In light of the facts, we were interested in preparing polyfluorinated oxadiazolyl pyridazinones (**Va–h**) to investigate whether these might show sufficiently enhanced chronic growth activity.

The next set of compounds, **VIa–c**, were prepared to study how the aromatic ring (R²) affects the activity by replacing it with the alkyl group and heterocycles. Finally, compounds **VIIa–f** were designed to probe the effect of replacing the *tert*-butyl group by the aliphatic chain and benzene ring.

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Table 1. Chronic Growth Activity of Compounds **IIIa–w**, **IVa–h**, **Va–h**, **Vla–c**, **Vlla–f**, and Azadirachtin on Armyworm *P. separata* (Walker) as well as EC₅₀ and PEC₅₀ Values

compd	X ¹	X ²	R ¹	R ²	EC ₅₀ (mg/L) (mean ± SE)	PEC ₅₀
IIIa	O	O	<i>tert</i> -butyl	C ₆ H ₅	12.2 ± 1.0	4.914
IIIb	O	O	<i>tert</i> -butyl	C ₆ H ₄ (4-NO ₂)	>300	<3.523
IIIc	O	O	<i>tert</i> -butyl	C ₆ H ₄ (3-CH ₃)	116.2 ± 4.9	3.935
IIId	O	O	<i>tert</i> -butyl	C ₆ H ₃ [3,5-(CH ₃) ₂]	>300	<3.523
IIIe	O	O	<i>tert</i> -butyl	C ₆ H ₄ (4-CH ₃)	19.9 ± 1.6	4.701
IIIf	O	O	<i>tert</i> -butyl	C ₆ H ₄ (4-F)	5.2 ± 0.6	5.284
IIIg	O	O	<i>tert</i> -butyl	C ₆ H ₄ (4-C ₂ H ₅)	14.5 ± 1.5	4.838
IIIh	O	O	<i>tert</i> -butyl	C ₆ H ₄ (4-Cl)	1.9 ± 0.6	5.721
IIIi	O	O	<i>tert</i> -butyl	C ₆ H ₄ (4-OCH ₃)	17.7 ± 0.5	4.752
IIIj	O	O	<i>tert</i> -butyl	C ₆ H ₄ (3-F)	7.1 ± 0.6	5.149
IIIk	O	O	<i>tert</i> -butyl	C ₆ H ₄ (2-F)	16.8 ± 2.8	4.775
IIIl	O	O	<i>tert</i> -butyl	C ₆ H ₂ (2,4-di-Cl-5-F)	23.1 ± 3.5	4.636
IIIm	S	O	<i>tert</i> -butyl	C ₆ H ₅	>300	<3.523
III n	S	O	<i>tert</i> -butyl	C ₆ H ₄ (4-F)	150.4 ± 15.3	3.822
IIIo	S	O	<i>tert</i> -butyl	C ₆ H ₄ (3-F)	243.4 ± 9.3	3.614
IIIp	S	O	<i>tert</i> -butyl	C ₆ H ₄ (2-F)	269.4 ± 11.3	3.569
IIIq	S	O	<i>tert</i> -butyl	C ₆ H ₄ (4-Cl)	180.4 ± 14.9	3.744
IIIr	S	O	<i>tert</i> -butyl	C ₆ H ₃ [3,5-(CH ₃) ₂]	>300	<3.523
III s	S	O	<i>tert</i> -butyl	C ₆ H ₄ (3-CH ₃)	>300	<3.523
III t	S	O	<i>tert</i> -butyl	C ₆ H ₄ (4-NO ₂)	>300	<3.523
IIIu	O	O	<i>tert</i> -butyl	C ₆ H ₄ (4-Br)	21.4 ± 1.7	4.669
IIIv	O	O	<i>tert</i> -butyl	C ₆ H ₃ (2,4-di-Cl)	27.8 ± 2.3	4.556
IIIw	O	O	<i>tert</i> -butyl	C ₆ H ₄ (2-Cl)	61.2 ± 3.8	4.213
IVa	O	S	<i>tert</i> -butyl	C ₆ H ₅	200.4 ± 9.6	3.698
IVb	O	S	<i>tert</i> -butyl	C ₆ H ₄ (4-F)	21.5 ± 2.3	4.667
IVc	O	S	<i>tert</i> -butyl	C ₆ H ₄ (2-F)	120.7 ± 7.8	3.918
IVd	O	S	<i>tert</i> -butyl	C ₆ H ₄ (4-Cl)	106.4 ± 5.7	3.973
IVe	O	S	<i>tert</i> -butyl	C ₆ H ₃ (2,4-di-Cl)	62.2 ± 3.3	4.206
IVf	O	S	<i>tert</i> -butyl	C ₆ H ₂ (2,4-di-Cl-5-F)	67.1 ± 4.2	4.173
IVg	O	S	<i>tert</i> -butyl	C ₆ H ₄ (3-CH ₃)	>300	<3.523
IVh	O	S	<i>tert</i> -butyl	C ₆ H ₃ [3,5-(CH ₃) ₂]	>300	<3.523
Va	O	O	<i>tert</i> -butyl	C ₆ H ₃ (2,6-di-F)	45.4 ± 2.8	4.343
Vb	O	O	<i>tert</i> -butyl	C ₆ H ₂ (2,4-di-Cl-3-NO ₂ -5-F)	>300	<3.523
Vc	O	O	<i>tert</i> -butyl	C ₆ (2,3,4,5,6-penta-F)	270.7 ± 8.8	3.567
Vd	O	O	<i>tert</i> -butyl	C ₆ H ₂ (2,4,5-tri-F)	75.7 ± 4.8	4.121
Ve	O	O	<i>tert</i> -butyl	C ₆ H (2,4,5-tri-F-3-CH ₃ O)	>300	<3.523
Vf	O	O	<i>tert</i> -butyl	C ₆ H ₃ (2-F-4-Cl)	10.4 ± 2.2	4.983
Vg	O	O	<i>tert</i> -butyl	C ₆ H (2,3,4,5-tetra-F)	240.6 ± 11.5	3.619
Vh	O	O	<i>tert</i> -butyl	C ₆ H ₄ (4-CF ₃)	34.1 ± 0.8	4.471
Vla	O	O	<i>tert</i> -butyl	CH ₃	>300	<3.523
Vlb	O	O	<i>tert</i> -butyl	2-furyl	>300	<3.523
Vlc	O	O	<i>tert</i> -butyl	2-thienyl	>300	<3.523
Vlla	O	O	C ₆ H ₅	C ₆ H ₅	91.5 ± 5.4	4.039
Vllb	O	O	<i>n</i> -butyl	C ₆ H ₅	>300	<3.523
Vllc	O	O	<i>n</i> -butyl	C ₆ H ₄ (4-F)	161.5 ± 7.6	3.792
Vlld	O	O	<i>n</i> -butyl	C ₆ H ₄ (4-Cl)	223.5 ± 13.6	3.651
Vlle	O	O	C ₂ H ₅	C ₆ H ₅	>300	<3.523
Vllf	O	O	C ₂ H ₅	C ₆ H ₄ (4-F)	>300	<3.523
azadirachtin					0.58 ± 0.05	6.237

The chronic growth inhibitory activity of the 48 synthesized compounds, including 20 earlier reported compounds (**IIIa–t**), was assessed using another insect, the armyworm, *Pseudaleia separata* (Walker). We also studied the quantitative structure–activity relationship of the recently and newly synthesized active compounds, and QSAR equations were established.

MATERIALS AND METHODS

Synthetic Procedures. All melting points (mp) were obtained with an electrothermal digital apparatus made in Shanghai and are uncorrected. ¹H NMR spectra were recorded on a Bruker WP-500SY (500 MHz) spectrometer with CDCl₃ as the solvent and TMS as the internal standard. Chemical shifts are reported in δ (parts per million) values. High-resolution mass spectra were recorded under electron impact (70 eV) condition using a MicroMass GCT CA 055 instrument. Combustion analyses for elemental composition were made with an Elementar vario EL III. Analytical thin-layer chromatography (TLC) was carried out on precoated plates (silica gel 60 F₂₅₄), and spots were visualized with ultraviolet (UV) light.

N-Chloroacetyl-*N'*-aroylhydrazines were prepared according to procedures found in the literature (14). 5-Aryl-2-(chloromethyl)-1,3,4-oxadiazoles (**III–v**) were prepared by the cyclodehydration of *N*-(chloroacetyl)-*N'*-aroylhydrazines in boiling POCl₃ (5). Syntheses of the target compounds, 2-alkyl-4-chloro-5-[5'-aryl-2'-(1',3',4'-oxa(thia)diazolyl)methoxy]-3(2*H*)-pyridazinones (**IIIu–w**, **IVa–h**, **Va–h**, **Vla–c**, and **Vlla–f**) were realized as described previously (5).

Synthetic Procedure for 5-Aryl-2-(chloromethyl)-1,3,4-thiadiazoles (IIIa–h). The mixture of the *N*-chloroacetyl-*N'*-aroylhydrazine (0.05 mmol), Lawesson reagent (0.05 mmol), and toluene (10 mL) was refluxed at 110 °C for 2 h under a nitrogen atmosphere. After cooling to room temperature, the mixture was treated with water (30 mL) and extracted with chloroform (3 × 10 mL). The organic layer was washed with water and dried (MgSO₄), and the solvent was removed under reduced pressure. The crude product was purified on a silica gel column eluted with petroleum ether (60–90 °C)/ethyl acetate (4:1) to give **IIIa–h**. The preparative and spectral data of **IIIa–v** are listed in **Table 2**.

Data for IIIu: yield, 72%; mp, 144–145 °C; ¹H NMR (CDCl₃) δ 1.65 [s, 9H, (CH₃)₃C], 5.55 (s, 2H, CH₂), 7.69 (d, 2H, ArH), 7.96 (d, 2H, ArH), 7.97 (s, 1H, PyH); EIMS, *m/z* (%) 438 (M⁺, 3), 383 (39), 373 (67), 347 (48), 237 (69), 183 (100), 155 (17), 57 (15). HRMS for C₁₇H₁₆N₄O₃ClBr: calcd, 438.0094; found, 438.0098.

Data for IIIv: yield, 81%; mp, 148–149 °C; ¹H NMR (CDCl₃) δ 1.65 [s, 9H, (CH₃)₃C], 5.58 (s, 2H, CH₂), 7.44 (dd, 1H, ArH), 7.61 (d, 1H, ArH), 7.99 (d, 1H, ArH), 7.96 (s, 1H, PyH); EIMS, *m/z* (%) 428 (M⁺, 5), 393 (14), 373 (67), 337 (64), 227 (72), 173 (100), 147 (24), 57 (7). HRMS for C₁₇H₁₅N₄O₃Cl₂: calcd, 428.0210; found, 428.0177.

Data for IIIw: yield, 63%; mp, 164–165 °C; ¹H NMR (CDCl₃) δ 1.68 [s, 9H, (CH₃)₃C], 5.60 (s, 2H, CH₂), 7.28–8.03 (m, 4H, ArH), 7.98 (s, 1H, PyH); EIMS, *m/z* (%) 394 (M⁺, 3), 359 (6), 339 (21), 303 (32), 193 (29), 139 (100), 111 (20), 57 (24). HRMS for C₁₇H₁₆-Cl₂N₄O₃: calcd, 394.0601; found, 394.0611.

Data for IVa: yield, 68%; mp, 145–146 °C; ¹H NMR (CDCl₃) δ 1.62 [s, 9H, (CH₃)₃C], 5.70 (s, 2H, CH₂), 7.50–8.10 (m, 5H, ArH), 7.90 (s, 1H, PyH); EIMS, *m/z* (%) 376 (0.5, M⁺), 341 (98), 284 (100), 175 (99), 121 (85). HRMS for C₁₇H₁₇ClN₄O₂S: calcd, 376.0761; found, 376.0772.

Data for IVb: yield, 71%; mp, 190–191 °C; ¹H NMR (CDCl₃) δ 1.60 [s, 9H, (CH₃)₃C], 5.82 (s, 2H, CH₂), 7.20–7.80 (m, 4H, ArH), 7.83 (s, 1H, PyH); EIMS, *m/z* (%) 394 (0.2, M⁺), 359 (78), 303 (47), 193 (100), 139 (82). HRMS for C₁₇H₁₆ClF₄N₄O₂S: calcd, 394.0667; found, 394.0674.

Data for IVc: yield, 62%; mp, 171–172 °C; ¹H NMR (CDCl₃) δ 1.60 [s, 9H, (CH₃)₃C], 5.81 (s, 2H, CH₂), 7.83 (s, 1H, PyH), 7.20–7.90 (m, 4H, ArH); EIMS, *m/z* (%) 394 (0.2, M⁺), 359 (100), 303 (55), 193 (90), 139 (90). HRMS for C₁₇H₁₆ClF₄N₄O₂S: calcd, 394.0667; found, 394.0661.

Data for IVd: yield, 65%; mp, 206–207 °C; ¹H NMR (CDCl₃) δ 1.62 [s, 9H, (CH₃)₃C], 5.70 (s, 2H, CH₂), 7.80 (s, 1H, PyH), 7.20–7.90 (m, 4H, ArH); EIMS, *m/z* (%) 410 (0.2, M⁺), 375 (88), 319 (72), 209 (100), 155 (88). HRMS for C₁₇H₁₆Cl₂N₄O₂S: calcd, 410.0371; found, 410.0374.

Data for IVe: yield, 66%; mp, 160–161 °C; ¹H NMR (CDCl₃) δ 1.60 [s, 9H, (CH₃)₃C], 5.84 (s, 2H, CH₂), 7.20–7.71 (m, 3H, ArH), 7.90 (s, 1H, PyH); EIMS, *m/z* (%) 444 (0.3, M⁺), 409 (87), 353 (83), 243 (100), 189 (98). HRMS for C₁₇H₁₅Cl₃N₄O₂S: calcd, 443.9981; found, 444.0001.

Data for IVf: yield, 60%; mp, 176–177 °C; ¹H NMR (CDCl₃) δ 1.61 [s, 9H, (CH₃)₃C], 5.82 (s, 2H, CH₂), 7.51–7.90 (m, 2H, ArH), 7.90 (s, 1H, PyH); EIMS, *m/z* (%) 462 (0.4, M⁺), 427 (81), 371 (86), 261 (97), 207 (100). HRMS for C₁₇H₁₄Cl₃FN₄O₂S: calcd, 461.9887; found, 461.9890.

Data for IVg: yield, 69%; mp, 126–127 °C; ¹H NMR (CDCl₃) δ 1.51 [s, 9H, (CH₃)₃C], 2.45 (s, 3H, CH₃), 5.70 (s, 2H, CH₂), 7.80 (s, 1H, PyH), 7.20–7.90 (m, 4H, ArH); EIMS, *m/z* (%) 390 (0.1, M⁺), 355 (77), 299 (79), 189 (81), 135 (100). HRMS for C₁₈H₁₉ClN₄O₂S: calcd, 390.0917; found, 390.0909.

Data for IVh: yield, 70%; mp, 148–149 °C; ¹H NMR (CDCl₃) δ 1.70 [s, 9H, (CH₃)₃C], 2.42 (s, 6H, CH₃), 5.71 (s, 2H, CH₂), 7.21–7.60 (m, 3H, ArH), 7.90 (s, 1H, PyH); EIMS, *m/z* (%) 404 (0.2, M⁺),

Table 2. Melting Point, Yield, and Mass Spectral Data of 5-Aryl-2-(chloromethyl)-1,3,4-oxa(thia)diazoles (**Ila–v**)

compd	X ²	R ²	mp (°C)	yield (%)	<i>m/z</i>
Ila	S	C ₆ H ₅	84–85	90	210 (M ⁺), 175, 135, 121, 103, 77
Ilb	S	C ₆ H ₄ (4-F)	118–119	82	228 (M ⁺), 193, 153, 139, 121, 95
Ilc	S	C ₆ H ₄ (2-F)	89–90	70	228 (M ⁺), 193, 153, 139, 121, 95
Ild	S	C ₆ H ₄ (4-Cl)	134–136	85	244 (M ⁺), 209, 169, 155, 137, 111
Ile	S	C ₆ H ₃ (2,4-di-Cl)	116–117	77	278 (M ⁺), 243, 203, 189, 171
Ilf	S	C ₆ H ₂ (2,4-di-Cl-5-F)	153–154	84	296 (M ⁺), 261, 221, 207, 189, 163
Ilg	S	C ₆ H ₄ (3-CH ₃)	82–83	80	224 (M ⁺), 189, 149, 135, 117, 91
Ilh	S	C ₆ H ₃ [3,5-(CH ₃) ₂]	92–93	75	238 (M ⁺), 203, 163, 149, 131, 105
Ili	O	C ₆ H ₃ (2,6-di-F)	137–139	81	228 (M ⁺), 193, 153, 139, 121, 95
Ilj	O	C ₆ H (2,4-di-Cl-3-NO ₂ -5-F)	177–179	78	210 (M ⁺), 175, 135, 121, 103, 77
Ilk	O	C ₆ (2,3,4,5,6-penta-F)	115–116	80	238 (M ⁺), 203, 163, 149, 131, 105
Ilm	O	C ₆ H ₂ (2,4,5-tri-F)	139–140	85	244 (M ⁺), 209, 169, 155, 137, 111
Ilm	O	C ₆ H (2,4,5-tri-F-3-CH ₃ O)	167–169	82	278 (M ⁺), 243, 203, 189, 171
Ilm	O	C ₆ H ₃ (2-F-4-Cl)	110–112	83	228 (M ⁺), 193, 153, 139, 121, 95
Ilo	O	C ₆ H (2,3,4,5-tetra-F)	155–156	80	296 (M ⁺), 261, 221, 207, 189, 163
Ilp	O	C ₆ H ₄ (4-CF ₃)	210–211	77	224 (M ⁺), 189, 149, 135, 117, 91
Iliq	O	CH ₃	135–137	70	132 (M ⁺), 83, 97, 43, 56, 15
Ilr	O	2-furyl	81–82	75	184 (M ⁺), 149, 135, 93, 95, 67
Ils	O	2-thienyl	163–165	78	200 (M ⁺), 166, 151, 109, 83, 111
Ilr	O	C ₆ H ₄ (4-Br)	187–188	70	274 (M ⁺), 237, 183, 155, 102
Ilu	O	C ₆ H ₃ (2,4-di-Cl)	108–110	80	262 (M ⁺), 213, 227, 173, 145, 109
Ilv	O	C ₆ H ₄ (2-Cl)	200–201	71	228 (M ⁺), 179, 193, 139, 111, 75

369 (98), 313 (69), 203 (92), 149 (100). HRMS for C₁₉H₂₁ClN₄O₂S: calcd, 404.1074; found, 404.1061.

Data for Va: yield, 63%; mp, 174–175 °C; ¹H NMR (CDCl₃) δ 1.65 [s, 9H, (CH₃)₃C], 5.60 (s, 2H, CH₂), 7.02 (t, *J* = 8.55 Hz, 2H, ArH), 7.56–7.59 (m, 1H, ArH), 7.96 (s, 1H, PyH); EIMS, *m/z* (%) 396 (M⁺, 3), 341 (57), 361 (9), 305 (51), 195 (42), 141 (100). HRMS for C₁₇H₁₅N₄O₃F₂Cl: calcd, 396.0801; found, 396.0781.

Data for Vb: yield, 57%; mp, 237–238 °C; ¹H NMR (CDCl₃) δ 1.57 [s, 9H, (CH₃)₃C], 5.92 (s, 2H, CH₂), 8.35 (s, 1H, PyH), 8.40 (d, *J* = 9.13 Hz, 1H, ArH); EIMS, *m/z* (%) 491 (M⁺, 3), 436 (100), 400 (32), 236 (37), 190 (13). HRMS for C₁₇H₁₃N₅O₃FCl: calcd, 490.9966; found, 490.9951.

Data for Vc: yield, 68%; mp, 191–193 °C; ¹H NMR (CDCl₃) δ 1.65 [s, 9H, (CH₃)₃C], 5.61 (s, 2H, CH₂), 7.97 (s, 1H, PyH); EIMS, *m/z* (%) 450 (M⁺, 4), 395 (100), 359 (81), 331 (5), 249 (45), 195 (97). HRMS for C₁₇H₁₂N₄O₃F₃Cl: calcd, 450.0518; found, 450.0498.

Data for Vd: yield, 72%; mp, 174–175 °C; ¹H NMR (CDCl₃) δ 1.65 [s, 9H, (CH₃)₃C], 5.60 (s, 2H, CH₂), 7.14–7.19 (m, 1H, Ar-H), 7.93–7.98 (m, 1H, Ar-H), 7.95 (s, 1H, Py-H); EIMS, *m/z* (%) 396 (M⁺, 3), 341 (57), 361 (9), 305 (51), 195 (42), 141 (100). HRMS for C₁₇H₁₅N₄O₃F₂Cl: calcd, 396.0801; found, 396.0781.

Data for Ve: yield, 63%; mp, 140–141 °C; ¹H NMR (CDCl₃) δ 1.65 [s, 9H, (CH₃)₃C], 4.12 (s, 3H, OCH₃), 5.57 (s, 2H, CH₂), 7.59–7.64 (m, 1H, ArH), 7.95 (s, 1H, PyH); EIMS, *m/z* (%) 396 (M⁺, 3), 341 (57), 361 (9), 305 (51), 195 (42), 141 (100). HRMS for C₁₇H₁₅N₄O₃F₂Cl: calcd, 396.0801; found, 396.0781.

Data for Vf: yield, 66%; mp, 119–120 °C; ¹H NMR (CDCl₃) δ 1.65 [s, 9H, (CH₃)₃C], 5.57 (s, 2H, CH₂), 7.27–7.35 (m, 2H, ArH), 7.96 (s, 1H, PyH), 8.04–8.07 (m, 1H, ArH); EIMS, *m/z* (%) 412 (M⁺, 3), 377 (9), 357 (53), 321 (50), 211 (63), 157 (100). HRMS for C₁₇H₁₅N₄O₃FCl₂: calcd, 412.0505; found, 412.0503.

Data for Vg: yield, 70%; mp, 186–187 °C; ¹H NMR (CDCl₃) δ 1.65 [s, 9H, (CH₃)₃C], 5.59 (s, 2H, CH₂), 7.77–7.79 (m, 1H, ArH), 7.94 (s, 1H, PyH); EIMS, *m/z* (%) 432 (M⁺, 6), 377 (100), 341 (44), 177 (93), 57 (18). HRMS for C₁₇H₁₃N₄O₃F₄Cl: calcd, 432.0612; found, 432.0616.

Data for Vh: yield, 64%; mp, 193–194 °C; ¹H NMR (CDCl₃) δ 1.65 [s, 9H, (CH₃)₃C], 5.58 (s, 2H, CH₂), 7.84 (d, *J* = 8.23 Hz, 2H, ArH), 7.97 (s, 1H, PyH), 8.23 (d, *J* = 8.23 Hz, 2H, ArH); EIMS, *m/z* (%) 428 (M⁺, 5), 373 (85), 337 (66), 173 (100), 145 (30). HRMS for C₁₈H₁₆N₄O₃F₃Cl: calcd, 428.0863; found, 428.0896.

Data for Via: yield, 57%; mp, 188–189 °C; ¹H NMR (CDCl₃) δ 1.57 [s, 9H, (CH₃)₃C], 2.53 (s, 3H, CH₃), 5.37 (s, 2H, CH₂), 7.85 (s, 1H, PyH); EIMS, *m/z* (%) 298 (M⁺, 7), 263 (17), 243 (100), 207 (92), 147 (57), 97 (17), 57 (13). HRMS for C₁₂H₁₅N₄O₃Cl: calcd, 298.0833; found, 298.0830.

Data for Vlb: yield, 70%; mp, 141–142 °C; ¹H NMR (CDCl₃) δ 1.65 [s, 9H, (CH₃)₃C], 5.53 (s, 2H, CH₂), 6.63–6.64 (m, 1H, furan-H), 7.26 (d, *J* = 3.81 Hz, 1H, furan-H), 7.69 (d, *J* = 1.35 Hz, 1H, furan-H), 7.95 (s, 1H, Py-H); EIMS, *m/z* (%) 350 (M⁺, 4), 315 (13), 295 (35), 259 (31), 149 (100), 95 (45), 57 (7). HRMS for C₁₅H₁₅N₄O₄-Cl: calcd, 350.0782; found, 350.0785.

Data for Vlc: yield, 75%; mp, 190–191 °C; ¹H NMR (CDCl₃) δ 1.65 [s, 9H, (CH₃)₃C], 5.53 (s, 2H, CH₂), 7.21–7.22 (m, 1H, thiophenyl-H), 7.63 (dd, *J* = 0.81 Hz, *J* = 0.81 Hz, 1H, thiophenyl-H), 7.69 (dd, *J* = 0.88 Hz, *J* = 0.88 Hz, 1H, thiophenyl-H), 7.98 (s, 1H, Py-H); EIMS, *m/z* (%) 366 (M⁺, 3), 331 (17), 311 (32), 275 (34), 165 (100), 111 (69), 57 (9). HRMS for C₁₅H₁₅N₄O₃S-Cl: calcd, 366.0553; found, 366.0591.

Data for Vlla: yield, 50%; mp, 177–178 °C; ¹H NMR (CDCl₃) δ 5.58 (s, 2H, CH₂), 7.39–7.62 (m, 5H, ArH), 7.56–8.12 (m, 5H, ArH), 8.03 (s, 1H, PyH); EIMS, *m/z* (%) 380 (M⁺, 4), 345 (18), 304 (36), 105 (100), 77 (20). Anal. Calcd (%) for C₁₉H₁₃ClN₄O₃: C, 59.93; H, 3.44; N, 14.71. Found: C, 59.73; H, 3.27; N, 14.84.

Data for Vllb: yield, 58%; mp, 143–145 °C; ¹H NMR (CDCl₃) δ 0.94 (t, *J* = 7.35 Hz, 3H, CH₃), 1.37–1.38 (m, 2H, CH₂), 1.77–1.79 (m, 2H, CH₂), 4.20 (t, *J* = 7.34 Hz, 2H, CH₂N), 5.57 (s, 2H, CH₂), 7.53–8.09 (m, 5H, ArH), 8.03 (s, 1H, PyH); EIMS, *m/z* (%) 360 (M⁺), 359 (6), 339 (21), 303 (32), 193 (29), 139 (100), 111 (20), 57 (24). Anal. Calcd (%) for C₁₇H₁₇ClN₄O₃: C, 56.59; H, 4.75; N, 15.52. Found: C, 56.72; H, 4.78; N, 15.73.

Data for Vllc: yield, 64%; mp, 140–142 °C; ¹H NMR (CDCl₃) δ 0.94 (t, *J* = 7.12 Hz, 3H, CH₃), 1.35–1.37 (m, 2H, CH₂), 1.77–1.78 (m, 2H, CH₂), 4.19 (t, *J* = 7.21 Hz, 2H, CH₂N), 5.58 (s, 2H, CH₂), 7.21–8.11 (m, 4H, ArH), 8.04 (s, 1H, PyH); EIMS, *m/z* (%) 378 (M⁺), 201 (12), 177 (8), 123 (100), 95 (32). Anal. Calcd (%) for C₁₇H₁₆-ClFN₄O₃: C, 53.90; H, 4.26; N, 14.79. Found: C, 53.81; H, 4.12; N, 14.49.

Data for Vlld: yield, 59%; mp, 168–170 °C; ¹H NMR (CDCl₃) δ 0.87 (t, *J* = 7.02 Hz, 3H, CH₃), 1.30–1.31 (m, 2H, CH₂), 1.70–1.72 (m, 2H, CH₂), 4.12 (t, *J* = 6.92 Hz, 2H, CH₂N), 5.49 (s, 2H, CH₂), 7.44–7.96 (m, 4H, ArH), 7.94 (s, 1H, PyH); EIMS, *m/z* (%) 395 (M⁺), 201 (20), 177 (8), 139 (100), 111 (32), 75 (16), 41 (18). Anal. Calcd (%) for C₁₇H₁₆Cl₂N₄O₃: C, 51.66; H, 4.08; N, 14.18. Found: C, 51.52; H, 3.88; N, 14.24.

Data for Vllle: yield, 66%; mp, 165–168 °C; ¹H NMR (CDCl₃) δ 1.37 (t, *J* = 7.23 Hz, 3H, CH₃), 4.27 (q, *J* = 7.22 Hz, 2H, CH₂N), 5.57 (s, 2H, CH₂), 7.53–8.09 (m, 5H, ArH), 8.04 (s, 1H, PyH); EIMS, *m/z* (%) 332 (M⁺), 297 (100), 194 (12), 159 (28), 105 (32), 103 (18). Anal. Calcd (%) for C₁₅H₁₃ClN₄O₃: C, 54.14; H, 3.94; N, 16.84. Found: C, 54.42; H, 3.88; N, 16.56.

Table 3. Reactions of Polyfluoro Aromatic Esters with Hydrazine Hydrate

R ²	reaction time (h)	reaction temp (°C)
C ₆ H (2,4-di-Cl-3-NO ₂ -5-F)	8	80–85
C ₆ H ₃ (2-F-4-Cl)		
C ₆ H ₄ (4-CF ₃)		
C ₆ H ₃ (2,6-di-F)	2–3	25–30
C ₆ H ₂ (2,4,5-tri-F)	4–5	0–5
C ₆ H (2,4,5-tri-F-3-CH ₃ O)		
C ₆ H (2,3,4,5-tetra-F)		
C ₆ (2,3,4,5,6-penta-F)	no desired product obtained	

Data for VIII: yield, 58%; mp, 195–197 °C; ¹H NMR (CDCl₃) δ 1.38 (t, *J* = 7.12 Hz, 3H, CH₃), 4.26 (q, *J* = 7.11 Hz, 2H, CH₂N), 5.56 (s, 2H, CH₂), 7.22–8.11 (m, 4H, ArH), 8.02 (s, 1H, PyH); EIMS, *m/z* (%) 350 (M⁺), 315 (14), 177 (18), 123 (100), 107 (11), 95 (45). Anal. Calcd (%) for C₁₅H₁₂ClF₃N₄O₃: C, 51.37; H, 3.45; N, 15.97. Found: C, 51.26; H, 3.49; N, 16.05.

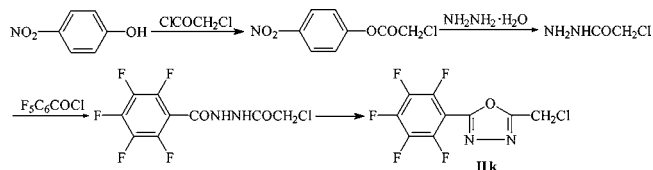
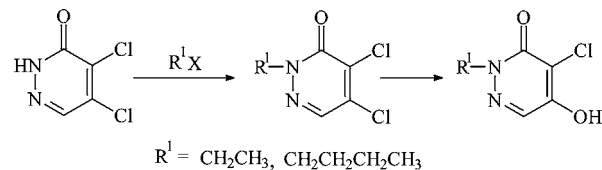
Insect. Larvae of the armyworm, *P. separata* (Walker), were reared continuously on freshly cut maize seedling under standard laboratory conditions (25 °C, 60% relative humidity, and a 16:8 h light/dark photoperiod).

Chronic Growth Bioassay. Effects of all 48 compounds and azadirachtin on larval growth were further assessed using second-instar larvae of the armyworm, *P. separata* (Walker). Fresh leaves containing the test substance and pure azadirachtin were prepared as described by Ismann (15, 16). Second-instar larvae of the armyworm (*n* = 30) were released on leaves with various concentrations of the analyzed compounds (0.01–500 mg/L). Three replicates were prepared for each concentration with 10 larvae in each replicate. Control diets were prepared in a similar way but with only solvent. After 5 days of exposure, all larvae were individually weighed to obtain EC₅₀ values (effective concentration to inhibit growth by 50% relative to controls). Data were mean values from triplicates [± standard error (SE)] and subjected to probit analysis. The data of EC₅₀ and the reciprocal logarithm of EC₅₀, pEC₅₀, are presented in **Table 1**.

RESULTS AND DISCUSSION

Synthesis. Starting from various aromatic esters, a set of aroylhydrazines, *N*-chloroacetyl-*N'*-aroylhydrazines and 5-aryl-2-chloromethyl-1,3,4-oxadiazoles, were synthesized following the procedure reported previously (5). With only a few exceptions, aroylhydrazines were synthesized by treatment of aromatic ester with excess hydrazine hydrate in ethanol at 80 °C for 7–8 h. However, attempts to obtain the polyfluoroaroylhydrazines under the above-mentioned experimental conditions were found to be unsuccessful due to hydrazino group substitution occurring on the benzene ring. Therefore, the aroylhydrazines containing polyfluorine atoms were prepared by modification of the conventional method. Conditions for hydrazinolysis of polyfluoro aromatic ester are given in **Table 3**.

2,3,4,5,6-Pentafluorobenzoylhydrazine could not be prepared directly by hydrazinolysis of ethyl 2,3,4,5,6-pentafluorobenzoate with hydrazine hydrate in ethanol. For this reason, 5-(2,3,4,5,6-pentafluorophenyl)-2-(chloromethyl)-1,3,4-oxadiazole (**IIIk**) could not be obtained by the general synthetic route. Thus, we adjusted our protocol and sought another practical approach for the synthesis of **IIIk** (17, 18). Heating 4-nitrophenol with chloroacetyl chloride at 40 °C in ethyl ether for 0.5 h led to chloroacetic acid 4-nitrophenyl ester, and subsequent treatment with hydrazine hydrate at 0 °C for 0.5 h in THF gave chloroacetylhydrazine. Further acylation of chloroacetylhydrazine with 2,3,4,5,6-pentafluorobenzoyl chloride followed by cyclodehydration in boiling POCl₃ afforded 5-(2,3,4,5,6-penta-

**Figure 2.** New synthetic route for 5-(2,3,4,5,6-pentafluorophenyl)-2-(chloromethyl)-1,3,4-oxadiazole.**Figure 3.** Synthetic route for 2-ethyl(or *n*-butyl)-4-chloro-5-hydroxy-3(2*H*)-pyridazinone.

fluorophenyl)-2-(chloromethyl)-1,3,4-oxadiazole (**IIIk**). The procedure is outlined in **Figure 2**.

Different from the synthesis of 2-*tert*-butyl-4-chloro-5-hydroxy-3(2*H*)-pyridazinone, which we described previously (5), 2-ethyl-4-chloro-5-hydroxy-3(2*H*)-pyridazinone and 2-*n*-butyl-4-chloro-5-hydroxy-3(2*H*)-pyridazinone are prepared by first alkylating 4,5-dichloro-3(2*H*)-pyridazinone followed by hydroxylation (19), as shown in **Figure 3**.

Chronic Growth Activity. Previously we reported the chronic growth activities against Asiatic corn borer, *Ostrinia furnacalis* (Guenee), of **IIIa–t** and also discussed the effects of the changes of gross structure on the chronic growth activity. In this study, we measured the chronic growth activity of 48 oxadiazolyl 3(2*H*)-pyridazinone derivatives having various substituents against the armyworm, *P. separata* (Walker).

Among the 48 compounds, compound **IIIh** was the most active. It was only slight weaker than azadirachtin. Compared to **IIIa**, with the introduction of F and Cl at the para position, increased activity, but a bulkier halogen (Br) and an electron-withdrawing group such as CF₃ or NO₂ decreased activity. Furthermore, the position of fluorine substituent in aryl derivatives (R² = aryl) played an important positive role in their potency in the order para > meta > ortho. Substitution by such groups as alkyl and alkoxy at meta and para positions somewhat reduced activity. The disubstituted analogues were found to be inferior to their respective unsubstituted (**IIIa**) and mono (para) substituted ones generally, except for the 2-F-4-Cl derivative (**VI**). Replacement of the oxo bridge by a mercapto bridge (**IIIm–t**) remarkably decreased the activity. These present results for *P. separata* are similar to those obtained earlier for *O. furnacalis*. The introduction of more than one fluorine atom on the benzene ring seems to be unfavorable to the activity to various degrees; the activity was found to fall off with increasing number of fluorine atoms. Thiadiazole compounds (**IVa–h**) exhibited lower activities compared to the corresponding oxadiazoles. The exchange of the *tert*-butyl group in the parent compounds (**IIIa**) for *n*-butyl (**VIIb,c**), ethyl (**VIIe,f**), or phenyl groups (**VIIa**) led to diminishing biological activity. These results indicate that the *tert*-butyl group on the nitrogen atom is very important for the insecticidal activity of the compounds. Loss of activity was also observed when the phenyl rings (R²) were replaced by methyl (**VIa**), furyl (**VIb**), or thienyl (**VIc**) groups. **Table 1** summarizes the insecticidal screening results of the studied compounds.

Physicochemical Parameters. To further explore the structural requirements of oxadiazolyl 3(2*H*)-pyridazinones for the activity, the activity values were quantitatively analyzed using

Table 4. Activity Observed (pEC₅₀) and Predicted and Residual Values for 19 Oxadiazolyl 3(2H)-Pyridazinones and Physicochemical Parameters for PLS Regression

compd	activity obsd (pEC ₅₀)	eq 2 PLS predicted	PLS residuals	dipole-mag	log P	MR
IIIa	4.914	4.821	0.093	6.07	2.95	50.605
IIIh	5.721	5.107	0.614	6.32	3.09	53.216
IIIi	5.284	5.264	0.020	6.34	2.69	50.729
IIIc	3.94	4.363	-0.428	5.72	3.02	53.979
IIIe	4.701	4.534	0.167	5.85	3.01	53.979
IIIg	4.838	4.830	0.008	6.22	3.52	54.612
IIIj	4.752	4.853	-0.101	6.24	3.52	54.202
IIIk	5.149	5.226	-0.077	6.31	2.69	50.729
IIIl	4.775	4.974	-0.199	6.17	2.89	50.729
IIIm	4.636	4.596	0.040	5.99	3.36	55.501
IIIu	4.669	4.917	-0.248	6.18	3.21	59.168
IIIv	4.556	4.292	0.264	5.78	3.52	59.826
IIIw	4.213	4.122	0.091	5.58	3.19	54.216
Va	4.343	4.327	0.016	5.63	2.75	50.853
Vc	3.567	3.451	0.116	5.21	3.06	12.168
Vd	4.121	4.183	-0.062	5.69	2.96	25.526
Vf	4.983	5.025	-0.042	6.29	3.23	54.601
Vg	3.619	3.684	-0.065	5.41	3.18	15.347
Vh	4.471	4.677	-0.206	6.15	3.66	52.571

physicochemical parameters and a regression method. All computations were done on a Silicon Graphics workstation running on the IRIX 6.5 operating system. Relevant computational modules were accessed from the Drug Discovery Workbench (DDW) of Cerius² (version 4.8). Molecular geometries were optimized under the universal force field (UFF) provided by Open Force Field (OFF) within Cerius².

The above qualitative structure-activity relationship analysis revealed that there were four possible regions governing the activity: the oxo bridge, the *tert*-butyl group, the oxadiazole ring, and the substituted phenyl. Therefore, particular attention was paid to those compounds, 2-*tert*-butyl-4-chloro-5-[5'-aryl-2'-(1',3',4'-oxadiazolyl)methoxy]-3(2H)-pyridazinones, and used for the regression analysis. Some compounds were excluded from the regression analysis because these compounds did not give acceptable EC₅₀ values. Nineteen 3(2H)-pyridazinones (X¹ = O, X² = O, R¹ = *tert*-butyl, R² = substituted phenyl) were used for quantitative analysis. The biological activities and physicochemical parameters of 19 compounds are summarized in **Table 4**.

To find out which of the parameters are important to activity, several equations were obtained according to different combinations of the parameters. The data matrix was analyzed by the partial least squares (PLS) method. The quality of each of the regression models was evaluated using the square of the correlation coefficient (*r*²); we examined only regressions with *r*² > 0.80. The most significant combination of physicochemical parameters affecting activity were dipole moment (*μ*), molar refractivity (MR), and log *P*.

The physical parameters dipole moment, molar refractivity, and log *P* are the independent variables of the regression, and the biological activity pEC₅₀ is the dependent variable. Finally, it was found that there was significant correlation between the pEC₅₀ and dipole moment and molar refractivity as showed by eq 1:

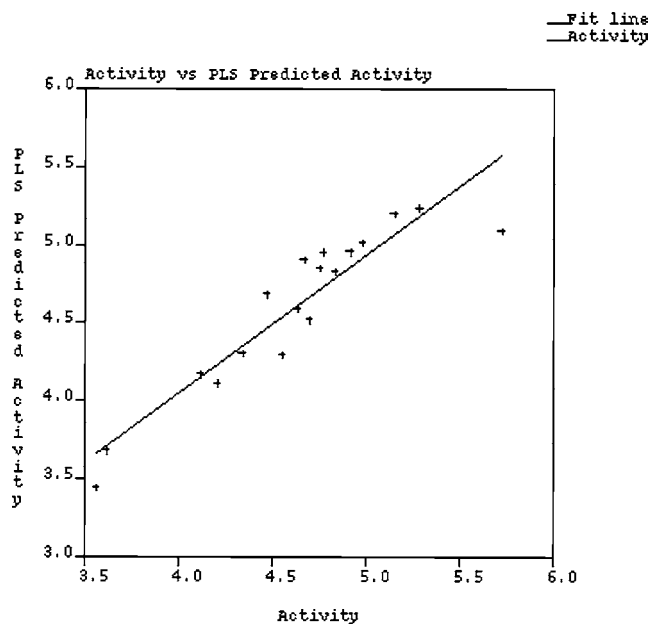
$$\text{pEC}_{50} = -3.49825 + 1.32928(\mu) + 0.0036254(\text{MR}) \quad (1)$$

$$r^2 = 0.803, \text{XV}r^2 = 0.684, \text{PRESS} = 1.687$$

In this and the following equations, important indices of goodness of fit are given: correlation coefficient, *r*²; cross-

Table 5. Correlation Matrix for Variables Used To Derive Equation 2

	dipole-mag	MR	log P	activity
dipole-mag	1.000			
MR	0.672	1.000		
log P	0.027	0.158	1.000	
activity	0.894	0.651	-0.151	1.000

**Figure 4.** Correlation between observed and estimated activities of 18 oxadiazolyl 3(2H)-pyridazinones using eq 2.

validation correlation coefficient, *XVr*²; and predicted sum of squares, PRESS. The cross-validation correlation coefficient *XVr*² is a measure of the predictive power of the equation. To investigate the need for a hydrophobic term in QSAR, we have used log *P* as an additional correlating parameter for obtaining a model for this set of 19 compounds. Addition of log *P* to the above model (eq 1) gave significant improvement in the statistics, yielding the following regression expression:

$$\text{pEC}_{50} = -2.2994 + 1.28423(\mu) + 0.0055719(\text{MR}) - 0.329534(\log P) \quad (2)$$

$$r^2 = 0.840, \text{XV}r^2 = 0.761, \text{PRESS} = 1.272$$

The coefficients of indicator parameters (dipole moment and MR) in eq 2 are positive. This indicates that the higher the dipole moment and MR values, the higher the activity. The negative coefficient of the log *P* term indicates that as molecular hydrophobicity (log *P*) of the present 19 compounds decreased, if substitution patterns are acceptable, their activity increased. The entire correlation matrix of the regression variables is shown in **Table 5**. The activities calculated by eq 2 are listed in **Table 4**. The correlation relationship between observed values and estimated values for eq 2 of the insecticidal activity is shown in **Figure 4**. The results indicate that log *P*, dipole moment, and MR values can be used to help predict the activity. Although a small set of analogues was used to develop these QSAR models, the models will provide insight in designing chronic growth activity oxadiazolyl 3(2H)-pyridazinones.

A relatively large number of the oxa(thia)diazolyl 3(2H)-pyridazinones have been prepared and characterized. Chronic growth activity (EC₅₀) against the armyworm, *P. separata* (Walker), was evaluated by comparison with the natural

antifeedant, azadirachtin. The results show that most of the oxa(thia)diazolyl 3(2H)-pyridazinones are able to inhibit armyworm growth to more or less extent depending on the structure of the compounds. Structure–activity relationship analysis of all of these oxadiazolyl 3(2H)-pyridazinones revealed that the $-OCH_2$ group, the *tert*-butyl group on the nitrogen atom, the oxadiazole ring, and the substituted benzene ring would be essential for the activity. Nineteen 2-*tert*-butyl-4-chloro-5-[5'-aryl-2'-(1',3',4'-oxadiazolyl)methoxy]-3(2H)-pyridazinones were subjected to a multiple linear regression analysis using the growth inhibition data pEC₅₀ values. Significant relationships were found between those three parameters and the activity against the armyworm. The chronic growth activities of oxadiazolyl 3(2H)-pyridazinones against the armyworm depend on the following factors: the dipole moment (μ), molar refractivity (MR), and log *P*. We hope that this model will be used in the future to design new effective antifeedants.

LITERATURE CITED

- Morimoto, M.; Kumeda, S.; Komai, K. Insect antifeedant flavonoids from *Gnaphalium affine* D. Don. *J. Agric. Food Chem.* **2000**, *48*, 1888–1891.
- Boyer, F. D.; Prangeb, T.; Ducrota, P. H. Synthesis of agarofuran antifeedants. Part 6: Enantioselective synthesis of a key decalinic intermediate. *Tetrahedron: Asymmetry* **2003**, *14*, 1153–1159.
- Verkerk, R. H. J.; Wright, D. J. Biological activity of neem seed kernel extracts and synthetic azadirachtin against larvae of *Plutella xylostella* L. *Pestic. Sci.* **1993**, *37*, 83–91.
- Ishihara, J.; Fukuzaki, T.; Murai, A. Synthetic studies on azadirachtin (part 3): asymmetric synthesis of the tricyclic dihydrofuran moiety of azadirachtin. *Tetrahedron Lett.* **1999**, *40*, 1907–1910.
- Cao, S.; Qian, X. H.; Song, G. H.; Chai, B.; Jiang, Z. S. Synthesis and antifeedant activity of new oxadiazolyl 3(2H)-pyridazinones. *J. Agric. Food Chem.* **2003**, *51*, 152–155.
- Hung, Q.; Qian, X.; Song, G.; Cao, S. The toxic and antifeedant activity of oxadiazole containing 2H-pyridazin-3-one derivatives against the armyworm, *Mythimna separata* Walker, and other insect as well as mite species. *Pest Manag. Sci.* **2003**, *59*, 933–939.
- Leyendecker, J.; Neubauer, H. J.; Kardorff, U.; Kuekenhoehner, T.; Kuenast, C.; Hofmeister, P. 3(2)-Pyridazinone derivatives and the use thereof for controlling pests. Ger. Offen. DE 3914337, 1990.
- Patani, G. A.; LaVoie, E. J. Bioisosterism: a rational approach in drug design. *Chem. Rev.* **1996**, *96*, 3147–3176.
- Ismail, F. M. D. Important fluorinated drugs in experimental and clinical use. *J. Fluorine Chem.* **2002**, *118*, 27–33.
- Lai, J. S.; Kool, E. T. Selective pairing of polyfluorinated DNA bases. *J. Am. Chem. Soc.* **2004**, *126*, 3040–3041.
- Abad, A.; Agulloa, C.; Cunat, A. C.; Jimenez, R.; Vilanova, C. Preparation and promotion of fruit growth in kiwifruit of fluorinated *N*-phenyl-*N'*-1,2,3-thiadiazol-5-yl ureas. *J. Agric. Food Chem.* **2004**, *52*, 4675–4683.
- Naumann, K. Research into fluorinated pyrethroid alcohols—an episode in the history of pyrethroid discovery. *Pestic. Sci.* **1998**, *52*, 3–20.
- Anderson, M.; Fisher, J. P.; Robinson, J.; Debray, P. H. Flufenoxuron—an acylurea acaricide/insecticide with novel properties. *Proc.—Br. Crop Prot. Conf.—Pests Dis.* **1986**, *1*, 89–96.
- Vakula, R. T.; Srinivasan, R. V. 1,3,4-Oxa(thia)diazoles: part VIII—synthesis and reactivity of 5-aryl-2-chloromethyl-1,3,4-oxadiazoles. *Indian J. Chem.* **1973**, *11*, 732–734.
- Isman, M. B. Growth inhibitory and antifeedant effects of azadirachtin on six noctuids of regional economic importance. *Pestic. Sci.* **1993**, *38*, 57–63.
- Isman, M. B.; Koul, O.; Luczynski, A.; Kaminski, J. Insecticidal and antifeedant bioactivities of neem oils and their relationship to azadirachtin content. *J. Agric. Food Chem.* **1990**, *38*, 1406–1411.
- Maki, A. K. T.; Yacouta-Nour, A.; Mostafa, M. M. Synthesis and characterization of new dioxouranium(VI) complexes of acid hydrazides. *J. Indian Chem. Soc.* **1995**, *72*, 447–451.
- Buyle, P. R. Sur less esters actives II. Preparation et hydrolyse basique des mono-, di- et trichlor-acetylhydrazides. *Helv. Chim. Acta* **1964**, *47*, 2449–2451.
- Kim, S.; Cho, S.; Kweon, D.; Lee, S.; Chung, J.; Shin, S.; Yoon, Y. Retro-ene reaction. II. Reaction of 4,5-dichloro-1-hydroxy-methylpyridazin-6-one with alkyl halides and carboxylic acid chlorides. *J. Heterocycl. Chem.* **1996**, *33*, 245–248.

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